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CONCERNING A FILING UNDER 35 U.S.C. 371

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U.S. APPLICATION NO. (If known, see 37 CFR 1.5)

INTERNATIONAL APPLICATION NO.
PCT/FR99/00643

INTERNATIONAL FILING DATE
March 19, 1999

PRIORITY DATE CLAIMED

TITLE OF INVENTION

VECTORS FOR TRANSERRING NUCLEIC ACIDS, COMPOSITIONS CONTAINING THEM

APPLICANT(S) FOR DO/EO/US
Ciolina, Carole et al.

AND THEIR USES

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
4. ☒ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. ☐ is transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☒ has been transmitted by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☒ A translation of the International Application into English (35 U.S.C. 371(c)(2)).
7. ☐ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
 - a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ have been transmitted by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☐ have not been made and will not be made.
8. ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)). *unexecuted
10. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11. to 16. below concern document(s) or information included:

11. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
12. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. ☐ A FIRST preliminary amendment.
☐ A SECOND or SUBSEQUENT preliminary amendment.
14. ☐ A substitute specification.
15. ☐ A change of power of attorney and/or address letter.
16. ☒ Other items or information: SEQUENCE LISTING W/DISKETTE AND STATEMENT

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17. ☒ The following fees are submitted:**BASIC NATIONAL FEE (37 CFR 1.492(a)(1)-(5)):**

Neither international preliminary examination fee (37 CFR 1.482)
nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO
and International Search Report not prepared by the EPO or JPO \$970.00

International preliminary examination fee (37 CFR 1.482) not paid to
USPTO but International Search Report prepared by the EPO or JPO. \$840.00

International preliminary examination fee (37 CFR 1.482) not paid to USPTO but
international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$690.00

International preliminary examination fee paid to USPTO (37 CFR 1.482)
but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$670.00

International preliminary examination fee paid to USPTO (37 CFR 1.482)
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CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE
Total claims	42 - 20 =	22	X \$18.00
Independent claims	2 - 3 =	--	X \$78.00

\$396.00

\$

MULTIPLE DEPENDENT CLAIM(S) (if applicable) + \$260.00

\$ 260.00

TOTAL OF ABOVE CALCULATIONS =

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\$ 130.00

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Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be
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NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:

CUSTOMER NO: 005487

SIGNATURE:

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NAME

34,647

REGISTRATION NUMBER

ST98008A US

1

VECTORS FOR TRANSFERRING NUCLEIC ACIDS, COMPOSITIONS
CONTAINING THEM AND THEIR USES

The present invention relates to new vectors
5 and to their use for transferring nucleic acids. More particularly, the invention relates to new vectors capable of directing nucleic acids towards cells or specific cellular compartments.

The transfer of nucleic acids is a technique
10 which forms the basis of all the major applications of biotechnology, and increasing the efficiency of transferring nucleic acids constitutes a very important challenge for the development of these applications. The efficiency of transferring nucleic acids depends on
15 numerous factors, among which are the capacity of the nucleic acids to cross the plasma membrane and their capacity to be transported within the cell up to the nucleus.

One of the major obstacles to the efficiency
20 of transferring nucleic acids stems from the fact that the genetic information is often poorly or not directed towards the target organ for which it is intended. Moreover, once the nucleic acid has penetrated into the target cell, it must still be directed towards the
25 nucleus so as to be expressed there. Furthermore, in the case of the transfer of nucleic acids into

differentiated or quiescent cells, the nucleus is delimited by a nuclear envelope which constitutes an additional barrier to the passage of these nucleic acids.

5 The recombinant viruses used as vectors possess sophisticated and efficient mechanisms for guiding the nucleic acids up to the nucleus. However, viral vectors have certain disadvantages inherent to their viral nature, which unfortunately cannot be
10 completed excluded. Another strategy consists either in transfecting naked DNA, or in using nonviral agents capable of promoting the transfer of DNA into eukaryotic cells. However, the nonviral vectors do not possess subcellular or nuclear targeting signals. Thus
15 the passage of naked DNA, or in combination with its nonviral agent, from the cytoplasm to the nucleus is, for example, a step which has a very low efficiency (Zabner et al., 1995).

 Various attempts to attach targeting signals
20 have thus been made. In particular, peptide fragments for targeting have been covalently attached to oligonucleotides in an antisense-oligonucleotide-targeting strategy [Eritja et al., *Synthesis of defined peptide-oligonucleotide hybrids containing a nuclear transport signal sequence*, Tetrahedron, Vol. 47, No. 24, pp. 4113-4120, 1991]. The complexes thus formed

are good candidates as potential inhibitors of expression of endogenous genes.

Transfection vectors comprising a synthetic polypeptide coupled by electrostatic interactions to a DNA sequence have also been described in patent application WO 95/31557, the said polypeptide consisting of a polymeric chain of basic amino acids, of an NLS peptide and of a hinge region which connects the NLS peptide to the polymeric chain and makes it possible to avoid steric interactions. However, this type of construct poses a problem of stability because the interactions called into play between the DNA and the targeting signal are of an electrostatic nature.

Moreover, nucleic acid-specific targeting peptide chimeras exist which are described in patent application WO 95/34664, the binding between the two being of a chemical nature. However, this method involves in particular enzymatic steps which are difficult to control and which do not make it possible to produce large quantities of nucleic acids.

Finally, it has been shown that it is possible to attach an NLS ("Nuclear Localization Signal") sequence to a plasmid DNA via a cyclopropylpyrroloindole (Nature Biotechnology, Volume 16, pp. 80-85, January 1998). However, a complete inhibition of the transcription of the gene of

interest because of the random attachment of several hundreds of NLS sequences onto the plasmid was observed. One solution proposed by the authors to remedy this consists in linking the NLS sequences onto linear fragments of DNA and then in coupling these modified fragments with other nonmodified fragments. However, this technique, like the preceding one, has the disadvantage of involving at least one enzymatic step.

Thus, all the methods proposed up until now do not make it possible to solve satisfactorily the difficulties linked to the targeting of double-stranded DNAs.

The present invention provides an advantageous solution to these problems. More particularly, the present invention uses oligonucleotides which are conjugated with targeting signals and which are capable of forming triple helices with one or more specific sequences present on a double-stranded DNA molecule.

Such a vector has the advantage of being able to direct a double-stranded DNA towards cells or specific cellular compartments by means of the targeting signal, without genetic expression being inhibited. The applicant has indeed shown that, by virtue of the formation of stable site-specific triple

helices, it is now possible to link a targeting signal to a double-stranded DNA in a site-specific manner. Consequently, it is possible to attach the targeting signal outside the expression cassette for the gene to be transferred. The applicant has thus shown that genetic expression in the cell is not inhibited in spite of the chemical modification of the DNA. Furthermore, the presence of a triple helix as a means for binding the targeting signal to the DNA is particularly advantageous because it makes it possible to preserve a DNA size which is appropriate for the transfection.

The vector obtained has, furthermore, the advantage of incorporating targeting signals which are very stably linked to the double-stranded DNA, in particular when the oligonucleotide capable of forming the triple helix is modified by the presence of an alkylating agent.

Another advantage of the invention is to make it possible to couple the DNA to be transferred to targeting signals whose number and nature are both controlled. Indeed, it is possible to control the number of targeting signals linked to each double-stranded DNA molecule by introducing a suitable number of specific sequences appropriate for the formation of triple helices into the said double-stranded DNA

molecule. Likewise, it is possible to introduce into the same double-stranded DNA molecule several oligonucleotides linked to different targeting signals (intracellular and/or extracellular), and in this case

5 it is also possible to determine beforehand the respective proportions thereof. In addition, these various targeting signals may be attached to the double-stranded DNA molecule in a manner stable to a greater or lesser degree depending on whether the

10 triple helix is formed with or without a covalent bond (that is to say with or without the use of an alkylating agent).

Finally, the functionalized triple helix obtained results only from chemical conversion steps

15 and may therefore be obtained in a simple manner, reproducibly and in very large, in particular industrial, quantities.

A first subject of the invention therefore relates to a vector useful in transfection, which is

20 capable of targeting a cell and/or a specific cellular compartment. More particularly, the vector according to the invention comprises a double-stranded DNA molecule and at least one oligonucleotide which is coupled to a targeting signal and which is capable of forming, by

25 hybridization, a triple helix with a specific sequence present on the said double-stranded DNA molecule.

For the purposes of the invention, "double-stranded DNA" is understood to mean a double-stranded deoxyribonucleic acid which may be of human, animal, plant, bacterial or viral origin, and the like. It may be obtained by any technique known to persons skilled in the art, and in particular by screening a library, by chemical or enzymatic synthesis of sequences obtained by screening libraries. It may be chemically or enzymatically modified.

10 This double-stranded DNA may be in linear or circular form. In the latter case, the double-stranded DNA may be in a supercoiled or relaxed state. Preferably, the DNA molecule is of circular form and in a supercoiled conformation.

15 The double-stranded DNA may also carry a replication origin which is functional or otherwise in the target cell, one or more marker genes, sequences for regulating transcription or replication, genes of therapeutic interest, anti-sense sequences which are modified or otherwise, regions for binding to other cellular components, and the like. Preferably, the double-stranded DNA comprises an expression cassette consisting of one or more genes of interest under the control of one or more promoters and of a
20 transcriptional terminator which are active in the target cells.

For the purposes of the invention,
"expression cassette for a gene of interest" is
understood to mean a DNA fragment which may be inserted
into a vector at specific restriction sites. The DNA
5 fragment comprises a nucleic acid sequence encoding an
RNA or a polypeptide of interest and comprises, in
addition, the sequences necessary for the expression
(enhancer(s), promoter(s), polyadenylation sequences
and the like) of the said sequence. The cassette and
10 the restriction sites are designed to ensure insertion
of the expression cassette into an open reading frame
appropriate for transcription and translation.

It is generally a plasmid or an episome
carrying one or more genes of therapeutic interest. By
15 way of example, there may cited the plasmids described
in Patent Applications WO 96/26270 and WO 97/10343
which are incorporated into the present by reference.

For the purposes of the invention, gene of
therapeutic interest is understood to mean in
20 particular any gene encoding a protein product having a
therapeutic effect. The therapeutic product thus
encoded may be in particular a protein or peptide. This
protein product may be homologous in relation to the
target cell (that is to say a product which is normally
25 expressed in the target cell when the latter has no
pathological condition). In this case, the expression

of a protein makes it possible, for example, to palliate an insufficient expression in the cell or the expression of a protein which is inactive or weakly active because of a modification, or to overexpress the said protein. The gene of therapeutic interest may also encode a mutant of a cellular protein, having increased stability, a modified activity, and the like. The protein product may also be heterologous in relation to the target cell. In this case, an expressed protein may, for example, supplement or provide an activity which is deficient in the cell, allowing it to combat a pathological condition, or to stimulate an immune response.

Among the therapeutic products for the purposes of the present invention, there may be mentioned more particularly enzymes, blood derivatives, hormones, lymphokines [interleukins, interferons, TNF, and the like (FR 92/03120)], growth factors, neurotransmitters or their precursors or synthesis enzymes, trophic factors [BDNF, CNTF, NGF, IGF, GMF, α FGF, β FGF, NT3, NT5, HARP/pleiotrophin, and the like, dystrophin or a minidystrophin (FR 91/11947)], the CFTR protein associated with cystic fibrosis, tumour suppressor genes [p53, Rb, Rap1A, DCC, k-rev, and the like (FR 93/04745)], the genes encoding factors involved in coagulation [factors VII, VIII, IX], the

genes involved in DNA repair, suicide genes [thymidine kinase, cytosine deaminase], the genes for haemoglobin or other protein carriers, the genes corresponding to the proteins involved in the metabolism of lipids, of
5 the apolipoprotein type chosen from apolipoproteins A-1, A-II, A-IV, B, C-I, C-II, C-III, D, E, F, G, H, J and apo(a), metabolic enzymes such as, for example, lipoprotein lipase, hepatic lipase, lecithin cholesterol acyl transferase, 7-alpha cholesterol
10 hydroxylase, phosphatidic acid phosphatase, or lipid transfer proteins such as cholesterol ester transfer protein and phospholipid transfer protein, an HDL-binding protein or a receptor chosen, for example, from the LDL receptors, the remnant chylomicron receptors
15 and the scavenger receptors, and the like.

The DNA of therapeutic interest may also be a gene or an anti-sense sequence, whose expression in the target cell makes it possible to control the expression of genes or the transcription of cellular mRNAs. Such
20 sequences can, for example, be transcribed in the target cell into RNAs which are complementary to cellular mRNAs and thus block their transcription to protein, according to the technique described in Patent EP 140 308. The genes of therapeutic interest also
25 comprise the sequences encoding ribozymes, which are capable of selectively destroying target RNAs (EP 321

201), or the sequences encoding single-chain intracellular antibodies such as, for example, ScFv.

As indicated above, the deoxyribonucleic acid may also comprise one or more genes encoding an
5 antigenic peptide, which is capable of generating an immune response in humans or in animals. In this specific embodiment, the invention therefore allows the production of vaccines or the carrying out of immunotherapeutic treatments applied to humans or to
10 animals, in particular against microorganisms, viruses or cancers. They may be in particular antigenic peptides specific for the Epstein-Barr virus, the HIV virus, the hepatitis B virus (EP 185 573), the pseudo-rabies virus, the syncytia forming virus, the influenza
15 virus, the cytomegalovirus (CMV), other viruses, or specific for tumours (EP 259 212).

Preferably, the deoxyribonucleic acid also comprises sequences allowing the expression of the gene of therapeutic interest and/or the gene encoding the
20 antigenic peptide in the desired cell or organ. They may be sequences which are naturally responsible for the expression of the gene considered when the sequences are capable of functioning in the infected cell. They may also be sequences of plant origin
25 (responsible for the expression of other proteins, or even synthetic). In particular, they may be promoter

sequences of eukaryotic or viral genes. For example, they may be promoter sequences derived from the genome of the cell which it is desired to infect. Likewise, they may be promoter sequences derived from the genome of the virus. In this regard, there may be mentioned, for example, the promoters of the E1A, MLP, CNV and RSV genes, and the like. In addition, these expression sequences may be modified by the addition of activating or regulatory sequences, and the like. The promoter may also be inducible or repressible.

A triple helix corresponds to the attachment of an oligonucleotide, modified or otherwise, to the double-stranded DNA by so-called "Hoogsteen" hydrogen bonds between the bases of the third strand and those of the region forming the double helix. These pairings occur in the large spiral of the double helix and are specific for the sequence considered [Frank-Kamenetski, M.D., *Triplex DNA Structures*, Ann. Rev. Biochem., 1995, 64, pp 65-95]. The specific sequence in the form of a double helix may be in particular a homopurine-homopyrimidine sequence. Two categories of triple helices can be distinguished according to the nature of the bases of the third strand [Sun, J. and C. Hélène, *Oligonucleotide-directed triple-helix formation*, Curr. Opin. Struct. Biol., 1993, 3, pp. 345-356]: the purine bases make it possible to obtain C-G⁺G and T-A⁺A

pairings, and the pyrimidine bases make it possible to obtain C-G⁺G⁺ and T-A⁺T pairings (the symbol ⁺ corresponds to a pairing with the third strand).

These structures have been characterized from
5 the physicochemical point of view by means of numerous
NMR (Nuclear Magnetic Resonance), hybridization
temperature or nuclease protection studies, which makes
it possible to define their properties and the
conditions for their stability. For the triple helices
10 with the third purine strand, this strand is
antiparallel relative to the purine strand of the DNA
and the formation of the triple helix highly depends on
the concentration of divalent ions: ions such as Mg²⁺
stabilize the structure formed with the third strand.
15 For the triple helices with a third homopyrimidine
strand, the latter is parallel relative to the purine
strand, and the formation of the triple helix is
dependent on the pH: an acidic pH of less than six
allows protonation of the cytosines and the formation
20 of an additional hydrogen bond stabilizing the triplet
C-G⁺C⁺. "Mixed" triple helices also exist for which the
third strand carries purine and pyrimidine bases. In
this case, the orientation of this third strand depends
on the base sequence of the homopurine region.
25 The oligonucleotides used in the present
invention are oligonucleotides which hybridize directly

with the double-stranded DNA. These oligonucleotides may contain the following bases:

- thymidine (T), which is capable of forming triplets with the doublets A.T of the double-stranded DNA (Rajagopal et al., Biochem 28 (1989) 7859);

- adenine (A), which is capable of forming triplets with the doublets A.T of the double-stranded DNA;

- guanine (G), which is capable of forming triplets with the doublets G.C of the double-stranded DNA;

- protonated cytosine (C⁺), which is capable of forming triplets with the doublets G.C of the double-stranded DNA (Rajagopal et al., cited above);

- uracil (U), which is capable of forming triplets with the base pairs A.U or A.T.

To allow the formation of a triple helix by hybridization, it is important that the oligonucleotide and the specific sequence present on the DNA are complementary. In this regard, in order to obtain the best attachment and the best selectivity, an oligonucleotide and a specific sequence which are perfectly complementary are used for the vector according to the invention. This may be in particular an oligonucleotide poly-CTT and a specific sequence poly-GAA. By way of example, there may be mentioned the

oligonucleotide having the sequence:

5'-GAGGCTTCTTCTTCTTCTTCTTCTT-3' (GAGG(CTT)₇, SEQ ID No.1), in which the bases GAGG do not form triple helices but make it possible to separate the

- 5 oligonucleotide from the coupling arm. The sequence (CTT)₇ (SEQ ID No.2) may also be mentioned. These oligonucleotides are capable of forming a triple helix with a specific sequence comprising complementary units (GAA). These may be in particular a region comprising
- 10 7, 14 or 17 GAA units. Another specific sequence of interest is the sequence: 5'-AAGGGAGGGAGGAGAGGAA-3' (SEQ ID No.3). This sequence forms a triple helix with the oligonucleotides: 5'-AAGGAGAGGGAGGGAGGAA-3' (SEQ ID No.4) or 5'-TTGGTGTGGTGGGTGGGT-3' (SEQ ID No.5).

- 15 In this case, the oligonucleotide binds in an orientation which is antiparallel to the polypurine strand. These triple helices are unstable in the presence of Mg²⁺ as is mentioned above (Vasquez et al., Biochemistry, 1995, 34, 7243-7251; Beal and Dervan,
- 20 Science, 1991, 251, 1360-1363).

- The specific sequence may be a sequence which is naturally present on the double-stranded DNA, or a synthetic sequence or a sequence of natural origin artificially introduced into it. It is particularly
- 25 advantageous to use an oligonucleotide capable of forming a triple helix with a sequence which is

naturally present on the double-stranded DNA. Indeed, this advantageously makes it possible to obtain the vectors according to the invention with unmodified plasmids, in particular commercial plasmids of the pUC, 5 pBR322 and pSV type, and the like. Among the natural homopurine-homopyrimidine sequences present in the double-stranded DNA, there may be mentioned a sequence comprising all or part of the sequence 5'-CTTCCCGAAGGGGAGAAAGG-3' (SEQ ID No.6) present in the 10 replication origin ColE1 of E. coli. In this case, the oligonucleotide forming the triple helix has the sequence: 5'-GAAGGGTTCTTCCCTCTTCC-3' (SEQ ID No.7) and binds alternately to the two strands of the double helix, as described by Beal and Dervan (J. Am. Chem. 15 Soc. 1992, 114, 4976-4982) and Jayasena and Johnston (Nucleic Acids Res. 1992, 20, 5279-5288). There may also be mentioned the sequence 5'-GAAAAAGGAAGAG-3' (SEQ ID No.8) of the β -lactamase gene of the plasmid pBR322 (Duval-Valentin et al., Proc. Natl. Acad. Sci. USA, 20 1922, 89, 504-508). Another sequence is AAGAAAAAAGAA (SEQ ID No. 9) present in the replication origin γ of the plasmids with a conditional replication origin such as pCOR.

Although perfectly complementary sequences 25 are preferred, it is understood, however, that certain mismatches may be tolerated between the sequence of the

oligonucleotide and the sequence present on the DNA, as long as they do not lead to an excessive loss of affinity. There may be mentioned the sequence 5'-AAAAAAGGGGAATAAGGG-3' (SEQ ID No. 10) present in the E. coli β -lactamase gene. In this case, the thymine interrupting the polypurine sequence may be recognized by a guanine of the third strand, thus forming a triplet ATG which is stable when it is surrounded by two TAT triplets (Kiessling et al., Biochemistry, 1992, 31, 2829-2834).

The oligonucleotide used may be natural (composed of natural bases which are unmodified or chemically modified). In particular, the oligonucleotide may advantageously exhibit certain chemical modifications which make it possible to increase its resistance or its protection in relation to nucleases, or its affinity in relation to the specific sequence or which also make it possible to provide other additional properties (J. Goodchild, *Conjugates of Oligonucleotides and Modified Oligonucleotides: A Review of their Synthesis and Properties*, Bioconjugate Chemistry, Vol. 1 No. 3, 1990, pp. 165-187).

According to the present invention, oligonucleotide is also understood to mean any succession of nucleosides having undergone a

modification of the backbone. Among the possible modifications, there may be mentioned oligonucleotide phosphorothioates which are capable of forming triple helices with DNA (Xodo et al., Nucleic Acids Res., 1994, 22, 3322-3330), likewise the oligonucleotides possessing formacetal or methylphosphonate backbones (Matteucci et al., J. Am. Chem. Soc., 1991, 113, 7766-7768). It is also possible to use the oligonucleotides synthesized with α -anomers of nucleotides, which also form triple helices with DNA (Le Doan et al., Nucleic Acids Res., 1987, 15, 7749-7760). Another modification of the backbone is the phosphoramidate bond. There may be mentioned, for example, the internucleotide bond N3'-P5' phosphoramidate described by Gryaznov and Chen, which gives oligonucleotides forming particularly stable triple helices with DNA (J. Am. Chem. Soc., 1994, 116, 3143-3144). Among the other modifications of the backbone, it is also possible to mention the use of ribonucleotides, of 2'-O-methylribose, of phosphotriester, and the like (Sun and Hélène, Curr. Opinion Struct. Biol., 116, 3143-3144). The phosphorus-containing backbone can finally be replaced with a polyamide backbone as in the PNAs (Peptide Nucleic Acid), which can also form triple helices (Nielson et al., Science, 1991, 254, 1497-1500; Kim et al., J. Am. Chem. Soc., 1993, 115, 6477-6481) or with a guanidine-

based backbone, as in the DNG (deoxyribonucleic guanidine, Proc. Natl. Acad. Sci. USA, 1995, 92, 6097-6101), polycationic analogues of DNA, which also form triple helices.

5 The thymine of the third strand may also be replaced with a 5-bromouracil, which increases the affinity of the oligonucleotide for DNA (Povsic and Dervan, L. Am. Chem. Soc., 1989, 111, 3059-3061). The third strand may also contain non natural bases among
10 which there may be mentioned 7-deaza-2'-deoxyxanthosine (Milligan et al., Nucleic Acids Res., 1993, 21, 327-333), 1-(2-deoxy- β -D-ribofuranosyl)-3-methyl-5-amino-1H-pyrazolo[4,3-d]pyrimidine-7-one (Koh and Dervan, J. Am. Chem. Soc., 1992, 114, 1470-1478), 8-oxoadenine, 2-
15 aminopurine, 2'-O-methylpseudoisocytidine, or any other modification known to persons skilled in the art (for a review see Sun and Hélène, Curr. Opinion Struct. Biol., 1993, 3, 345-356).

 The object of another type of modification of
20 the oligonucleotide is more particularly to improve the interaction and/or the affinity between the oligonucleotide and the specific sequence. In particular, a completely advantageous modification consists in coupling an alkylating agent to the
25 oligonucleotide. The binding may take place either chemically or photochemically by means of a

photoreactive functional group. Advantageous alkylating agents are in particular photoactivable alkylating agents, for example psoralens. Under the action of light, they form covalent bonds at the level of the pyrimidine bases of the DNA. When these molecules are intercalated at the level of the 5'-ApT-3' or 5'-TpA-3' sequences in a double-stranded DNA fragment, they form bonds with both strands. This light-induced binding reaction can occur at a specific site of the plasmid.

As has been highlighted previously, one advantage of the present invention is therefore the possibility of forming very stable and site-specific triple helices between the oligonucleotide and a specific sequence of the double-stranded DNA by means of a covalent bond formed via an alkylating agent.

The length of the oligonucleotide used in the method of the invention is at least 3 bases, and preferably between 5 and 30 bases. An oligonucleotide having a length of between 10 and 30 bases is advantageously used. The length may of course be adapted on a case-by-case basis by persons skilled in the art according to the desired selectivity and stability of the interaction.

The oligonucleotides according to the invention may be synthesized by any known technique. In particular, they may be prepared by means of nucleic

acid synthesizers. Any other method known to persons skilled in the art may quite obviously be used.

For the purposes of the invention, "targeting signal" is understood to mean targeting molecules of a varied nature. It represents in most cases peptides known for targeting. They may be used to interact with a component of the extracellular matrix, a plasma membrane receptor, in order to target an intracellular compartment or in order to improve the intracellular flow of DNA, during the nonviral transfer of genes in gene therapy.

These targeting signals may comprise, for example, growth factors (EGF, PDGF, TGF β , NGF, IGF, I, FGF), cytokines (IL-1, IL-2, TNF, Interferon, CSF), hormones (insulin, growth hormone, prolactin, glucagon, thyroid hormone, steroid hormones), sugars which recognize lectins, immunoglobulins, ScFv's, transferrin, lipoproteins, vitamins such as vitamin B12, peptide or neuropeptide hormones (tachykinins, neurotensin, VIP, endothelin, CGRP, CCK, and the like), or any unit recognized by the integrins, for example the peptide RGD, or by other extrinsic proteins of the cell membrane.

It is also possible to use whole proteins, or peptide sequences derived from these proteins, or alternatively peptides which bind to their receptor and

which are obtained by the "phage display" technique or by combinatory synthesis.

Intercellular targeting signals may also be envisaged. Many nuclear homing sequences (NLS) of varied amino acid compositions have been identified and make it possible to get different proteins involved in the nuclear transport of proteins or of nucleic acids. Among these are in particular short sequences (the NLS of the SV40 T antigen (PKKKRKV, SEQ ID No. 11) is one example), bipartite sequences (the NLS of nucleoplasmin which contains two essential domains for nuclear transport: KRPAATKKAGQAKKKLKD, SEQ ID No. 12), or the M9 sequence (NQSSNFGPMKGGNFGGRSSGPYGGGGQYFAKPRNQGGY, SEQ ID No. 13) of the hnRNP A1 protein. The proteins carrying these NLS sequences bind to specific receptors, such as the receptors of the importin or karyopherin family, for example. The role of these sequences is to direct the DNA inside the nucleus where it is then immediately available for the transcription machinery, and may be expressed.

The "mixed" targeting signals, that is to say which can serve both for intracellular and extracellular targeting, also come within the scope of the present invention. It is possible to mention, for example, the sugars which target lectins which are present on the cell membrane but also at the level of

the nuclear pores. Targeting by these sugars therefore also relates both to extracellular targeting and nuclear import.

Other signals are involved in mitochondrial targeting (for example the N-terminal part of rat ornithine transcarbamylase (OTC) allows the targeting of mitochondria) or in homing onto the endoplasmic reticulum. Finally, some signals allow nuclear retention or retention at the level of the endoplasmic reticulum (such as the sequence KDEL).

Advantageously, the targeting signals according to the invention make it possible to direct the double-stranded DNA specifically towards certain cells or certain cellular compartments. By way of example, the targeting signals according to the invention can target receptors or ligands at the cell surface, in particular the receptors for insulin, for transferrin, for folic acid, or any other growth factor, cytokines, or vitamins, or particular polysaccharides at the surface of the cell or on the neighbouring extracellular matrix.

The synthesis of oligonucleotide-targeting signal chimaeras occurs on a solid phase or in solution and takes into account the very different stability properties of the oligonucleotides and targeting signals (Erijita, R. et al., *Synthesis of defined*

peptide-oligonucleotide hybrids containing a nuclear transport signal sequence, Tetrahedron, 1991, 47(24), pp. 4113-4120]. In solution, it is possible to envisage couplings in one step: the targeting signal may, for example, be synthesized with a group carrying disulphide, maleimide, amine, carboxyl, ester, epoxide, cyanogen bromide or aldehyde functional groups, and may be coupled to an oligonucleotide modified by a thiol, amine or carboxyl terminal group at the 3' or 5' position. These couplings are made by establishing disulphide, thioether, ester, amide or amine bonds between the oligonucleotide and the targeting signal. Any other method known to persons skilled in the art may be used, such as bifunctional coupling reagents, for example.

Another subject of the invention relates to compositions comprising a vector as defined above.

Advantageously, the vectors according to the invention may also be combined with one or more agents known for transfecting DNA. There may be mentioned by way of example the cationic lipids which possess advantageous properties. These vectors actually consist of a cationic polar part which interacts with the DNA, and a hydrophobic lipid part which promotes cellular penetration and renders the ionic interaction with DNA insensitive to the external medium. Specific examples

of cationic lipids are in particular the monocationic lipids (DOTMA: Lipofectin⁷), some cationic detergents (DDAB), lipopolyamines and in particular dioctadecylamidoglycyl spermine (DOGS) or

5 5-carboxyspermylamide of palmitoylphosphatidyl-ethanolamine (DPPES), whose preparation has been described, for example, in patent application EP 394 111. Another advantageous lipopolyamine family is represented by the compounds described in patent

10 application WO 97/18185 incorporated into the present by way of reference. Numerous other cationic lipids have been developed and may be used with the vectors according to the invention.

Among the synthetic transfection agents

15 developed, the cationic polymers of the polylysine and the DEAE-dextran type are also advantageous. It is also possible to use the polyethylenimine (PEI) and polypropylenimine (PPI) polymers which are commercially available and may be prepared according to the method

20 described in patent application WO 96/02655.

In general, any synthetic agent known to transfect nucleic acid may be combined with the vectors according to the invention.

The compositions may, in addition, comprise

25 adjuvants capable of combining with the vector according to the invention/transfection agent complexes

and of improving the transfecting power thereof. In another embodiment, the present invention therefore relates to compositions comprising a vector as defined above, one or more transfection agents as defined above and one or more adjuvants capable of combining with the vector according to the invention/transfection agent(s) complexes and of improving the transfecting power thereof. The presence of this type of adjuvant (lipids, peptides or proteins, for example) can advantageously make it possible to increase the transfecting power of the compounds.

In this regard, the compositions of the invention may comprise, as adjuvant, one or more neutral lipids whose use is particularly advantageous. The applicant has indeed shown that the addition of a neutral lipid makes it possible to enhance the formation of nucleolipid particles and to promote the penetration of the particle into the cell by destabilizing its membrane.

More preferably, the neutral lipids used within the framework of the present invention are lipids containing 2 fatty chains. In a particularly advantageous manner, natural or synthetic lipids which are zwitterionic or lacking ionic charge under physiological conditions are used. They may be chosen more particularly from dioleoylphosphatidylethanolamine

(DOPE), oleoylpalmitoylphosphatidylethanolamine (POPE), di-stearoyl, -palmitoyl, -myristoyl phosphatidylethanolamines as well as their derivatives which are N-methylated 1 to 3 times, phosphatidyl-
5 glycerols, diacylglycerols, glycosyldiacylglycerols, cerebrosides (such as in particular galactocerebrosides), sphingolipids (such as in particular sphingomyelins) or asialogangliosides (such as in particular asialoGM1 and GM2).

10 These different lipids may be obtained either by synthesis or by extraction from organs (for example the brain) or from eggs, by conventional techniques well known to persons skilled in the art. In particular, the extraction of the neutral lipids may be
15 carried out by means of organic solvents (see also Lehninger, Biochemistry).

 The applicant has demonstrated that it was also particularly advantageous to use, as adjuvant, a compound directly involved or otherwise in the
20 condensation of the DNA (WO 96/25508). The presence of such a compound in the composition according to the invention makes it possible to reduce the quantity of transfecting compound, with the beneficial consequences resulting therefrom from the toxicological point of
25 view, without any damaging effect on the transfecting activity. Compound involved in the condensation of the

nucleic acid is intended to define a compound which compacts, directly or otherwise, the DNA. More precisely, this compound may either act directly at the level of the DNA to be transfected, or may be involved
5 at the level of an additional compound which is directly involved in the condensation of this nucleic acid. Preferably, it acts directly at the level of the DNA. In particular, this agent which is involved in the condensation of the DNA may be any polycation, for
10 example polylysine. According to a preferred embodiment, this agent may also be any compound which is derived, as a whole or in part, from a histone, a nucleolin, a protamine and/or from one of their derivatives, as a whole or in part, of peptide units
15 (KTPKKAKKP SEQ ID No.16) and/or (ATPAKKAA SEQ ID No. 17), it being possible for the number of units to vary between 2 and 10. In the structure of the compound according to the invention, these units may be repeated continuously or otherwise. They may thus be separated
20 by linkages of a biochemical nature, for example one or more amino acids, or of a chemical nature.

The subject of the invention is also the use of the vectors as defined above for manufacturing a medicament intended to treat diseases by transfection
25 of DNA into primary cells or into established cell lines. They may be fibroblast cells, muscle cells,

nerve cells (neurons, astrocytes, glial cells), hepatic cells, haematopoietic cell line (lymphocytes, CD34, dendritic cells, and the like), epithelial cells and the like, in differentiated or pluripotent form

5 (precursors).

The vectors of the invention may, by way of illustration, be used for the *in vitro*, *ex vivo* or *in vivo* transfection of DNA encoding proteins or polypeptides.

10 For usage *in vivo*, either in therapy or for studying the regulation of the genes or the creation of animal models of pathological conditions, the compositions according to the invention can be formulated for administration by the topical, cutaneous, oral,

15 rectal, vaginal, parenteral, intranasal, intravenous, intramuscular, subcutaneous, intraocular, transdermal, intratracheal or intraperitoneal route, and the like. Preferably, the compositions of the invention contain a vehicle which is pharmaceutically acceptable for an

20 injectable formulation, in particular a direct injection into the desired organ, or for administration by the topical route (on the skin and/or the mucous membrane). They may be in particular isotonic sterile solutions, or dry, in particular freeze-dried,

25 compositions which, upon addition, depending on the case, of sterilized water or of physiological saline,

allow the constitution of injectable solutions. The nucleic acid doses used for the injection as well as the number of administrations may be adapted according to various parameters, in particular according to the mode of administration used, the relevant pathological condition, the gene to be expressed, or the desired duration of treatment. As regards more particularly the mode of administration, it may be either a direct injection into the tissues or the circulatory system, or a treatment of cells in culture followed by their reimplantation *in vivo* by injection or transplantation.

The invention relates, in addition, to a method of transfection of DNA into cells comprising the following steps:

- (1) synthesis of the oligonucleotide-targeting signal chimera according to the method described above,
 - (2) bringing the chimera synthesized in (1) into contact with a double-stranded DNA so as to form triple helices,
 - (3) optionally, complexing the vector obtained in (2) with one or more transfection agents and/or one more adjuvants, and
 - (4) bringing the cells into contact with the complex formed in (2) or, if applicable, in (3).
- The cells may be brought into contact with the complex by incubating the cells with the said

complex (for uses *in vitro* or *ex vivo*), or by injecting the complex into an organism (for uses *in vivo*).

The present invention thus provides a particularly advantageous method for the treatment of diseases by administration of a vector according to the invention containing a nucleic acid capable of correcting the said disease. More particularly, this method is applicable to diseases resulting from a deficiency in a protein or peptide product, the administered DNA encoding the said protein or peptide product. The present invention extends to any use of a vector according to the invention for the *in vivo*, *ex vivo* or *in vitro* transfection of cells.

The invention also relates to any recombinant cell containing a vector as defined above. It involves preferably eukaryotic cells, for example yeast, animal cells, and the like. These cells are obtained by any technique allowing the introduction of a DNA into a given cell known to persons skilled in the art.

The following examples which are intended to illustrate the invention without limiting its scope make it possible to demonstrate other characteristics and advantages of the present invention.

25 **FIGURES**

Figure 1: Coupling of an oligonucleotide (ODN) and the

peptide maleimide-NLS.

Figure 2: Analysis, on a 15% polyacrylamide gel, of the oligonucleotide-peptide (Pso-GA₁₉-NLS) chimera by proteolytic action of trypsin.

5 1 = oligo Pso-GA₁₉-SH

2 = oligonucleotide-peptide Pso-GA₁₉-NLS chimera

3 = oligonucleotide-peptide Pso-GA₁₉-NLS chimera after digestion with trypsin

Figure 3: schematic representation of the plasmid
10 pXL2813.

Figure 4: schematic representation of the plasmid pXL2652.

Figure 5: analysis on a 15% polyacrylamide gel of the formation of triple helices between the plasmid pXL2813
15 and the oligonucleotide-peptide Pso-GA₁₉-NLS chimera.

The oligonucleotide pso-GA₁₉-NLS and the plasmid are mixed in a buffer containing 100 mM MgCl₂. The molar excess of oligonucleotide relative to the plasmid varies from 0 to 200. The mixture is photoactivated,

20 after leaving overnight at 37°C, and then digested with two restriction enzymes which cut the plasmid on either side of the region of formation of the triple helices.

1 = no oligonucleotide

2 = molar excess of oligonucleotide relative to the
25 plasmid of 15

3 = molar excess of oligonucleotide relative to the

plasmid of 50

4 = molar excess of oligonucleotide relative to the
plasmid of 100

5 = molar excess of oligonucleotide relative to the
5 plasmid of 200

6 = photoactivated oligonucleotide alone

7 = nonphotoactivated oligonucleotide alone

8 = molar excess of oligonucleotide relative to the
plasmid of 30, the mixture not having been

10 photoactivated.

Figure 6: Expression *in vitro* of the transgene (β -galactosidase) for tests carried out with the plasmid pXL2813 alone, the vector pXL2813-Pso-GA₁₉-NLS, and without plasmid.

15 Figure 7: Characterization of the peptide part of the oligonucleotide-peptide Pso-GA₁₉-NLS chimera by interaction with importin 60-GST (analysis on a 15% polyacrylamide gel).

1 = oligo Pso-GA₁₉ (1 μ g)

20 2 = oligonucleotide-peptide Pso-GA₁₉-NLS chimera (1 μ g)

3 = supernatant recovered after incubation of glutathione-beads coated with importins 60 and the oligonucleotide Pso-GA₁₉, and separation of the pellet of beads (containing the components which interact with

25 the importins) from the supernatant

4 = pellet recovered after incubation of glutathione-

beads coated with importins 60 and the oligonucleotide Pso-GA₁₉, and separation of the pellet of beads (containing the components which interact with the importins) from the supernatant

- 5 5 = supernatant recovered after incubation of the glutathione-beads coated with importins 60 and the oligonucleotide-peptide Pso-GA₁₉-NLS chimera, and separation of the pellet of beads (containing the components which react with the importins) from the
- 10 supernatant
- 6 = pellet recovered after incubation of the glutathione-beads coated with importins 60 and the oligonucleotide-peptide Pso-GA₁₉-NLS chimera, and separation of the pellet of beads (containing the
- 15 components which interact with the importins) from the supernatant.

Figure 8: Analysis on a 15% polyacrylamide gel of the oligonucleotide-peptide (GA₁₉-NLS) chimera by the proteolytic action of trypsin.

- 20 1 = oligo GA₁₉-SH (200 ng)
2 = oligonucleotide-peptide GA₁₉-NLS chimera (1μg) before purification by high-performance liquid chromatography
3 = oligonucleotide-peptide GA₁₉-NLS chimera (1μg) after
- 25 purification
4 = oligonucleotide-peptide GA₁₉-NLS chimera after

digestion with trypsin (1 μ g).

Figure 9: Graphical representation of the kinetics of formation of the triple helices (% of triple helix sites occupied as a function of time) between the plasmid pXL2813 and the chimera GA₁₉-NLS.

Figure 10: Characterization of the peptide part of the oligonucleotide-peptide GA₁₉-NLS chimera by interaction with the importin 60-GST.

- 1 = oligonucleotide-peptide GA₁₉-NLS chimera,
- 10 2 = oligo GA₁₉,
- 3 = supernatant recovered after incubation of the glutathione-beads coated with importins 60 and the oligonucleotide-peptide GA₁₉-NLS chimera, and separation of the pellet of beads (containing the components which
- 15 interact with the importins) from the supernatant,
- 4 = pellet recovered after incubation of the glutathione-beads coated with importins 60 and the oligonucleotide-peptide GA₁₉-NLS chimera, and separation of the pellet of beads (containing the components which
- 20 interact with the importins) from the supernatant,
- 5 = supernatant recovered after incubation of the glutathione-beads coated with importins 60 and the oligonucleotide-peptide GA₁₉ and separation of the pellet of beads (containing the components which
- 25 interact with the importins) from the supernatant,
- 6 = pellet recovered after incubation of the

glutathione-beads coated with importins 60 and the oligonucleotide GA₁₉, and separation of the pellet of beads (containing the components which interact with the importins) from the supernatant.

5 Figure 11: Schematic representation of the plasmid pXL2997.

Figure 12: Graphical representation of the kinetics of formation of the triple helices (% of triple helix sites occupied as a function of time) between the
10 plasmid pXL2997 and the chimera pim-NLS.

Figure 13: Histogram representing the β -galactosidase activity *in vivo* in human pulmonary tumours H1299 of the plasmids pXL2813 (indicated Bgal in the figure) and pXL2813-Pso-GA₁₉-NLS (indicated NLS-Bgal in the figure)
15 in RLU ("Relative Light Unit") per tumour.

The transfection was performed using the electrotransfer techniques as described in applications WO 99/01157 and WO 99/01158.

20 MATERIALS AND METHODS

1. Coupling of the oligonucleotides and the peptides

Oligonucleotides

The oligonucleotides used are the sequence
25 5'-AAGGAGAGGAGGGAGGGAA-3' (SEQ ID No. 4) 19 bases long and referenced under the name "GA₁₉" in the text which

follows, or the sequence 5'-GGGGAGGGGGAGG-3' (SEQ ID No. 15) 13 bases long and referenced under the name "pim" in the text which follows (because it is the sequence of the protooncogene pim-1).

5 The oligonucleotides noted GA₁₉-SH or pim-SH have the same sequences as GA₁₉ and pim, respectively, and a thiol group at the 5' end, with a spacer of six carbon between the thiol and the phosphate of the 5' end. The oligonucleotides noted Pso-GA₁₉-SH have a thiol
10 group at the 3' (SH) end, and in addition a psoralen at the 5' (Pso) end, with a spacer of six carbons between the psoralen and the phosphate of the 5' end. The oligonucleotides noted Pso-GA₁₉ do not have a thiol group.

15 The nomenclature used is summarized in Table I below:

name of the oligonucleotide	modification of the 3' end	modification of the 5' end
GA ₁₉	none	none
pim	none	none
GA ₁₉ -SH	none	thiol group
pim-SH	none	thiol group
Psi-GA ₁₉	none	psoralen
Pso-GA ₁₉ -SH	thiol group	psoralen

Table 1

These freeze-dried oligonucleotides are taken up in a 100 mM trimethylammonium acetate buffer, pH = 7.

5 Peptides

The peptides used for the couplings are synthesized by an automatic machine in the solid phase. They contain:

- either the nuclear localization sequence of the SV40
- 10 T antigen (PKKKRKV, SEQ ID No. 11),
- or the same sequence, mutated (PKKKRKV, SEQ ID
- No. 14) which allows neither targeting nor nuclear
- transport (because of the mutation).

- These peptides also carry a spacer of four
- 15 amino acids at the N-terminal end: KGAG. The N-terminal lysine is chemically modified: it contains a maleimide group and the ϵ carbon and a protecting group
 - 9-fluorenylmethyloxycarbonyl (Fmoc) on the amine of the α carbon. This Fmoc group absorbs at 260 nm, which
 - 20 makes it possible to monitor the peptide by reversed-phase high-performance liquid chromatography. The C-terminal group is also protected (CONH₂ group), the protection being added at the end of peptide synthesis.

The representation of the peptides is

25 indicated in Table II below:

name of the peptide	sequence and modifications
maleimide-NLS	maleimide-KGAGPKKKRKV-CONH ₂ Fmoc
maleimide-NLSmutated	maleimide-KGAGPKNKRKV-CONH ₂ Fmoc

Table II

The freeze-dried peptides are picked up in a 100 mM triethylammonium acetate buffer, pH = 7. The concentration is 0.4 mg/ml.

5

Couplings

For the coupling with the oligonucleotides, the strategy used consists in reacting the thiol group carried by the oligonucleotide and the maleimide group carried by the peptide [Eritja, R., et al., *Synthesis of defined peptide-oligonucleotide hybrids containing a nuclear transport signal sequence*, Tetrahedron, 1991, **47**(24), pp. 4113-4120].

The oligonucleotide is added to the peptide solution in an equimolar quantity and the reaction medium is left for 2 hours at room temperature. The oligonucleotide-peptide chimera is purified by reversed-phase high-pressure liquid chromatography on a Vydac C8 column containing spheroidal silica having a diameter of 5 μm and a porosity of 300 Å. Use is made

of a 0.1 M triethylammonium acetate (TEAA) buffer and an acetonitrile gradient passing from 5% to 50% over 35 minutes. The products are detected at 260 nm.

5 Analysis of the chimeras by digestion with trypsin

The oligonucleotide-peptide conjugates are subjected to the proteolytic action of trypsin which makes it possible to reveal the peptide part of the chimera [Reed, M.W. et al., *Synthesis and evaluation of*
10 *nuclear targeting peptide-anti-sense oligodeoxynucleotide conjugates*, Bioconjugate Chemistry, 1995, 6, pp.101-108]. Solutions containing 1 µg of oligonucleotide-peptide in 7 µl of 0.1 M triethylammonium (TEAA) purification buffer are mixed
15 with 1 µl of a trypsin solution (5 mg/ml). 1 µl of 100 mM Tris-HCl buffer, pH = 9, and 1 µl of 500 mM EDTA are added thereto. After digesting for one hour, the samples are deposited into the wells of a 15% polyacrylamide, 7 M urea, gel. The electrophoresis is
20 performed in 100 mM Tris buffer, pH 8.3 containing 90 mM boric acid and 1 mM EDTA. The nucleic acids are visualized by silver staining using a Biorad kit.

25 2. Formation of triple helices with the chimeras

Plasmid

The plasmid used to study the formation of triple helices with the GA₁₉-peptide chimeras is called pXL2813 (7257 pb, cf Figure 3). This plasmid expresses the β -galactosidase gene under the control of the strong promoter of the cytomegalovirus (CMV) early genes, as well as the ampicillin resistance gene.

Upstream of the promoter, between positions 7238 and 7256, the GA₁₉ sequence (5'-AAGGAGAGGAGGGAGGGAA-3', SEQ ID No. 4) was cloned according to conventional molecular biological techniques.

It was cloned into the plasmid pXL2652 (of 7391 bp and which is schematically represented in Figure 4) which expresses the β -galactosidase gene under the control of the strong promoter of the cytomegalovirus (CMV) early genes, as well as the ampicillin resistance gene. This promoter is derived from the pCDNA3, the LacZ and its polyA are derived from the pCH110, and the remainder is derived from pGL2.

The sequences were cloned upstream of the promoter between the unique cleavage sites for the enzymes MunI and XmaI. For that, the two complementary oligonucleotides containing the sequence to be cloned

6651 (5'-AATGATTCTCTCTCCTCCCTCCCTTAC-3' SEQ ID No.18)
and 6652 (3'-CTAAGGAGAGGAGGGGAGGGAATGGG-5' SEQ ID No.19)
were heated for 5 minutes at 95°C and then hybridized
by allowing the temperature to fall slowly. The plasmid
5 pXL2652 was then digested with the enzymes MunI and
XmaI for 2 hours at 37°C and the products of this
double digestion were separated by electrophoresis on a
1% agarose gel followed by staining with ethidium
bromide.

10 The fragment of interest for the remainder of
the cloning was eluted according to the Jetsorb
protocol (Genomed) and 200 ng of this fragment were
linked to 10 ng of the mixture of oligonucleotides
hybridized by T4 ligase, for 16 hours at 16°C.

15 Competent E. coli bacteria of the DH5 α strain
were transformed by eletroporation with the reaction
product, plated on Petri dishes containing LB medium
and ampicillin. The ampicillin-resistant clones were
selected and the DNA was extracted by alkaline lysis
20 and analysed on a 1% agarose gel. A clone corresponding
in size to the expected product was sequenced.

In the case where the oligonucleotide is pim,
the plasmid used to study priple helix formation is
pXL2997 represented in Figure 11. This plasmid
25 expresses the gene for β -galactosidase under the
control of the strong promoter of the Cytomegalovirus

(CMV) early genes, as well as the gene for resistance to Ampicillin.

Upstream of the promoter, the sequence pim (SEQ ID No. 15) was cloned according to conventional molecular biology methods.

Formation of the triple helices

The formation of the triple helices is performed by mixing the plasmid pXL2813 (3 pmol of plasmid, that is to say 15 μ g) or pXL2997, depending on the case, and variable quantities of oligonucleotides or of chimeras in a 100 mM Tris-HCl buffer, pH = 7.5, containing 100 mM MgCl₂.

15 Photoactivation of the triple helices

After incubating overnight, the mixture is irradiated, in ice, for 15 minutes, using a monochromatic lamp having a wavelength of 365 nm (Biorad). The product of photoactivation is digested with the restriction enzymes MfeI and SpeI and analysed on a 15% polyacrylamide, 7 M urea, gel with 100 mM Tris migration buffer, pH = 8.3, containing 90 mM boric acid and 1 mM EDTA. The nucleic acids are visualized by silver staining using a Biorad kit [Musso, M., J.C. Wang, and M.W.V. Dyke, *In vivo persistence of DNA triple helices containing psoralen-conjugated*

oligodeoxyribonucleotides, Nucleic Acid Research, 1996, 24(24), pp. 4924-4932].

Method of studying the triple helices

- 5 This method is based on the principle of
extrusion chromatography of solutions containing the
plasmid and the oligonucleotide which are capable of
forming a triple helix. The exclusion columns used
(Columns, Linkers 6, Boehringer Mannheim) are composed
10 of Sepharose beads and have an exclusion limit of
194 base pairs, which makes it possible to retain in
the column the oligonucleotides not paired with the
plasmids.
- In a first instance, the oligonucleotides are
15 radioactively labelled at their 3' end with Terminal
Transferase with the aid of [α -³⁵S]dATP. The protocol
used is from Amersham: 10 pmol of oligonucleotides are
incubated for 2 hours at 37°C, with 50 μ Ci of
[α -³⁵S]dATP in the presence of 10 units of Terminal
20 Transferase, in a volume of 50 μ l of buffer containing
sodium cacodylate. The percentage of labelled
oligonucleotides is evaluated according to the
following method: 1 μ l of a sample, diluted 1/100, of
the solution after labelling is deposited on Whatman
25 DE81 paper, in duplicate. One of the two papers is
washed twice for 5 minutes with 2HSSC, for 30 seconds

with water and for 2 minutes with ethanol. The radioactivities of the two papers are compared. The radioactivity of the washed paper corresponds to the ^{35}S effectively incorporated.

5 The formation of the triple helices occurs in a volume of 35 μl . The concentrations of plasmids (40 nM) and of oligonucleotides (20 nM), the buffer used (100 mM Tris-HCl pH = 7.5, 50 mM of MgCl_2) and the temperature (37°C) are fixed whereas the incubation
10 time varies from 1 to 24 hours. The Sepharose columns are equilibrated, before use, with the reaction buffer and centrifuged at 2200 rpm for 4 minutes, in order to cause them to settle. 25 μl of the reaction medium are deposited on the columns and the latter are centrifuged
15 under the same conditions as above. 25 μl of the reaction buffer are then deposited on the columns which are again centrifuged. The eluate is recovered.

 The radioactivity contained in 5 μl of the reaction medium, noted cpm(deposit), and that contained
20 in the eluate, noted cpm(eluate), are evaluated, which makes it possible to estimate the percentage of oligonucleotides eluted:

$$\% \text{ of oligonucleotides eluted} = \text{cpm}(\text{eluate}) / [5 \text{ H cpm}(\text{deposit})] \text{ H } 100.$$

25 The percentage of plasmids which are effectively eluted during the experiments is evaluated

by estimating the optical density at 260 nm of the eluate and that of the deposit, which makes it possible to calculate the percentage of oligonucleotides attached to the total number of plasmids.

5 % of oligonucleotides attached = [% oligonucleotides eluted/% plasmids eluted] H 100.

This parameter makes it possible to evaluate the percentage of sites of formation of triple helices (there is one per plasmid pXL2813 or pXL2997) which are
10 effectively occupied, taking into account the concentrations of plasmids (noted [plasmid]) and of oligonucleotides (noted [oligo]) used during the reaction of formation of the triple helices:
% triple helix sites occupied = % of oligonucleotides
15 attached H [oligo]/[plasmid].

3. Interaction with the importins

Recombinant proteins

20 The importin 60 subunit used to study the interaction with the oligonucleotide-peptide (NLS or NLSmutated) conjugates is of murine origin and is fused with Glutathione S-Transferase (GST). The sequence of importin 60 was cloned into a vector pGEX-2T in order
25 to fuse it with GST. The recombinant protein was produced in Escherichia coli [Imamoto, N. et al., In

vivo evidence for involvement of a 58kDa component of nucleopore-targeting complex in nuclear protein import, The EMBO Journal, 1995, 14(15), pp. 3617-3626].

5 Interactions with the recombinant proteins

All the interaction experiments are carried out in the binding buffer (20 mM HEPES, pH = 6.8, 150 mM potassium acetate, 2 mM magnesium acetate, 2 mM DTT and 100 μ g/ml BSA)

- 10 In a first instance, the recombinant proteins are incubated in the presence of Sepharose beads coated with glutathion (Pharmacia Biotech), 1 μ g of recombinant protein is used for 10 μ l of beads. After incubating for 30 minutes at room temperature in 500 μ l
15 of binding buffer, the mixture is centrifuged at 2000 g for 30 seconds, and the supernatant is removed. The beads are washed five times by resuspending in 500 μ l of binding buffer followed by centrifugation, as described above. The beads are resuspended in binding
20 buffer in order to obtain a suspension containing 50% of beads coated with recombinant proteins.

- In a second instance, 60 μ l of the suspension containing 50% of beads coated with recombinant proteins are incubated with 2 μ g of oligonucleotide or
25 of oligonucleotide-peptide, in a volume of 500 μ l of binding buffer. After incubating for 30 minutes at room

temperature, the mixture is centrifuged at 2000 g for 30 seconds, and the supernatant is removed. 30 μ l of the supernatant are collected in order to analyse the fraction not attached to the beads. The beads are washed five times by resuspending in 500 μ l of binding buffer followed by centrifugation, as described above. The beads are resuspended in 15 μ l of loading buffer (0.05% bromophenol blue, 40% sucrose, 0.1 M EDTA, pH = 8, 0.5% sodium lauryl sulphate) and heated for 10 minutes at 90°C. The content of the supernatant and of the pellet is analysed on a 15% polyacrylamide, 7 M urea, gel, with a 100 mM Tris migration buffer, pH 8.3, containing 90 mM boric acid and 1 mM EDTA. The nucleic acids are visualized by silver staining using a Biorad kit [Rexach, M. and G. Blobel, *Protein import into nuclei: association and dissociation reactions involving transport substrate, transport factors, and nucleoporins*, Cell, 1995, 83, pp. 683-692].

20 4. Transfection of cells

Cell culture

The cell type used is NIH 3T3 (ATCC CRL-1658). It consists of mouse fibroblasts. These cells are cultured in a modified Dulbecco's medium, with 25 4.5 g/l glucose (DMEM - Gibco), 2 mM glutamine, penicillin (100 U/ml) and streptomycin (100 μ g/ml), and

10% foetal calf serum (Gibco). They are incubated at 37°C in an incubator containing 5% CO₂.

Transfection

- 5 One day before the transfection, the wells of a 24-well plate are inoculated with 50,000 cells per well. The vectors are diluted in 150 mM NaCl and mixed with a cationic lipid (the compound having the condensed formula
- 10 H₂N(CH₂)₃NH(CH₂)₄NH(CH₂)₃NHCH₂COglyN[(CH₂)₁₇CH₃]₂ described in patent application WO 97/18185 under the number (6)) diluted in 150 mM NaCl. The mixture is made with 6 nmol of lipid per microgram of plasmid. This mixture is diluted 1/10 in serum-free culture medium and deposited
- 15 on the cells. After incubating at 37°C in an incubator containing 5% CO₂ for two hours, 10% foetal calf serum is added.

Quantification of the β-galactosidase

- 20 After incubating for 48 hours, the cells are washed twice with PBS and lysed with 250 μl of lysis buffer (Promega). The β-galactosidase is quantified according to the protocol "Lumigal β-Galactosidase genetic reporter system" (Clontech). The activity is
- 25 measured on a Lumat LB9501 luminometer (Berthold). The quantity of proteins is measured with the BCA kit

(Pierce).

Examples

This example illustrates the possibility of
5 coupling the oligonucleotide Pso-GA₁₉-SH to the peptide
maleimide-NLS.

The oligonucleotide Pso-GA₁₉-SH, of sequence
5'-AAGGAGAGGAGGGAGGAA-3', SEQ ID No.4) with a thiol
group at the 3' end, was coupled to the peptide NLS
10 which carries a maleimide group at its N-terminal end
according to the method described above in the
"Materials and Methods" part under the section
"coupling of the oligonucleotides and the peptides".

The coupling is monitored by reversed-phase
15 high-performance liquid chromatography (HPLC). It
occurs with an equimolar stoichiometry: in two hours,
the coupling is complete and the chimera is purified by
HPLC.

The reaction scheme for the coupling is
20 represented in Figure 1.

The oligonucleotide-peptide (Pso-GA₁₉-NLS)
chimera was analysed by polyacrylamide gel
electrophoresis after proteolytic action of trypsin
which makes it possible to reveal the presence of the
25 peptide part of the chimera, after migration on a
denaturing polyacrylamide gel and silver staining of

the nucleic acids (as indicated in the "Materials and Methods" part under the section "Analysis of the chimeras by digestion with trypsin").

The chimera Pso-GA₁₉-NLS exhibits a retarded electrophoretic migration compared with the oligonucleotide Pso-GA₁₉-SH, and the product of proteolytic digestion is visualized at an intermediate level between the levels of migration of Pso-GA₁₉-NLS and of Pso-GA₁₉-SH, as shown in Figure 2.

The chimera Pso-GA₁₉-NLS therefore contains a peptide part which is accessible to trypsin. These results clearly show that the coupling between the oligonucleotide and the peptide occurred, and the coupling yield is high.

Example 2

This example illustrates the formation of triple helices between the plasmid pXL2813 and the chimera Pso-GA₁₉-NLS which is modified by a photoactivable alkylating agent. This example also indicates the proportion of plasmids modified according to the molar excess of oligonucleotides relative to the plasmid.

The plasmid pXL2813, represented in Figure 3, comprises the homopurine sequence complementary to GA₁₉ which is capable of forming a triple helix with the

oligonucleotides GA₁₉, Pso-GA₁₉, Pso-GA₁₉-SH or Pso-GA₁₉-NLS. Divalent cations such as Mg²⁺ stabilize these triple helices. The oligonucleotide Pso-GA₁₉-NLS and the plasmid are mixed in a buffer containing 100 mM of MgCl₂. The molar excess of oligonucleotide relative to the plasmid varies from 0 to 200.

After incubating for 12 hours at 37°C, the mixture is photoactivated for 15 minutes (as indicated in the "Materials and Methods" part under the section "Photoactivation of the triple helices"), and then digested with the two restriction enzymes MfeI and SpeI which cut the plasmid on either side of the region of formation of the triple helices. A nucleic acid fragment is thus released which contains 70 base pairs after digestion of the nonmodified plasmid pXL2813. The fragment obtained with the plasmid and the oligonucleotide forming a triple helix covalently bound to the double helix has 70 base pairs plus 19 bases of the oligonucleotide. By migration on a denaturing polyacrylamide gel, it is thus possible to separate these fragments of different size: the fragments derived from the plasmids modified by a triple helix have a shorter migration distance than the fragments derived from nonmodified plasmids. It is therefore possible, by electrophoresis on a denaturing polyacrylamide gel, to quantify the proportion of

modified plasmids according to the molar excess of oligonucleotide relative to the plasmid.

The results are indicated in Figure 5. For a molar excess of oligonucleotide relative to the plasmid greater than 50, all the plasmids are modified and are combined with an oligonucleotide Pso-GA₁₉-NLS. Without photoactivation, the retardation of the digestion fragment is lost in a denaturing gel.

It thus appears that the triple helices formed with the oligonucleotides Pso-GA₁₉-SH or Pso-GA₁₉-NLS are therefore indeed covalently bound to the double helix after photoactivation.

Moreover, by digesting the remainder of the plasmid backbone and by analysing as above, it is possible to check that the photoactivation does not result in a nonspecific covalent binding of the oligonucleotide Pso-GA₁₉-NLS outside the region containing the sequence capable of forming a triple helix.

These results clearly demonstrate the conditions which make it possible to specifically and covalently combine a peptide sequence with a plasmid, and is outside the promoter regulating the expression of the transgene, which prevents the expression of the transgene from being affected.

Example 3

This example illustrates the capacity of the transgene to be expressed *in vitro*, although the plasmid is modified.

- 5 The expression of β -galactosidase by the plasmids pXL2813 which are nonmodified or which are combined with an oligonucleotide Pso-GA₁₉-NLS was thus compared by transfection of NIH3T3 cells.

- 10 The results are reported in Figure 6, and the mean values obtained (∇ standard deviation) are assembled in the following Table III:

plasmid	expression of the transgene (in RLU/ μ g of protein)
pXL2813	253759 ∇ 48545
pXL2813-Pso-GA ₁₉ -NLS	473984 ∇ 111476
without plasmid	129 ∇ 85

Table III

- 15 It can be observed that the expression of the transgene increases following the modification provided by the covalent attachment of the triple helix upstream of the promoter.

 These results clearly demonstrate that the formation of the triple helices does not affect the

expression of the transgene *in vitro*. In contrast, by virtue of the nuclear homing of the plasmid because of its coupling with the targeting sequence NLS, more plasmids reached the nucleus, resulting in an increase in the transfection efficiency.

Example 4

This example is intended to verify that the *in vivo* expression is not inhibited by the presence of a targeting signal associated with the plasmid via a triple helix.

The experiment consists in transfecting pXL2813 and pXL2813-Pso-GA₁₉-NLS into human pulmonary tumours H1299 using the electrotransfer techniques described in applications WO 99/01157 and WO 99/01158. The experiment was carried out in 18 to 20 g female nude mice.

The mice are monolaterally implanted with 20 mm³ H1299 tumour grafts. The tumours develop, reaching a volume of 200 to 300 mm³. The mice are separated according to the size of their tumours and divided into homogeneous groups. The mice are then anaesthetized with a Ketamine/Xylazine mixture (commercially available). The plasmid solution (8 µg of DNA/40 µl of 150 mM sodium chloride) is longitudinally injected at the periphery of the tumour with the aid of

a Hamilton syringe.

The lateral faces of the tumour are covered with conducting gel and the tumour is placed between two stainless steel flat electrodes 0.45 to 0.7 cm apart. The electrical pulses are applied with the aid of a commercial square pulse generator (Electro-pulsateur PS 15, Jouan, France) 20 to 30 seconds after the injection. An oscilloscope makes it possible to control the intensity in volts, the duration in 10 milliseconds and the frequency in hertz of the pulses which should be delivered at 500 V/cm for 20 milliseconds at 1 hertz.

For the evaluation of the tumour transfection, 10 and 7 mice respectively for the 15 plasmid pXL2813 and pXL2818-Pso-GA₁₉-NLS are humanely killed 2 days after the injection of the plasmid. The tumours are collected, weighed and ground in a lysis buffer (Promega) supplemented with antiproteases (Complete, Boehringer). The suspension obtained is 20 centrifuged in order to obtain a clear supernatant. After having incubated 10 μ l of this supernatant with 250 μ l of reaction buffer (Clontech) for 1 hour in the dark, the β -galactosidase is measured with the aid of a commercial luminometer. The results are expressed in 25 total RLU ("Relative Light Unit") per tumour.

It is observed that the plasmid pXL2813-Pso-

GA₁₉-NLS expresses the transgene *in vivo* at a level greater than or equal to that obtained with the unmodified plasmid pXL2813 (see Figure 13).

Example 5

5 This example illustrates the characterization of the peptide part of the Pso-GA₁₉-NLS conjugates, that is to say the verification of the targeting properties of the NLS peptide combined with the constructs according to the invention.

10 The peptide sequence used, the NLS signal of the SV40 T antigen, is recognized by receptors of the α -karyopherin family. The murine equivalent, called importin 60, fused with a glutathione S-transferase group, was used to characterize the oligonucleotide-
15 peptide conjugates. It was performed according to the method described in the "Materials and Methods" part under the section "Interaction with the importins".

 The interactions between the glutathion-beads coated with importins 60 and the oligonucleotides GA₁₉-
20 NLS or Pso-GA₁₉-NLS were studied. After incubating for 30 minutes at room temperature, the pellet of beads (containing the components which interact with the importins) is separated from the supernatant.

 The result of this characterization is
25 reported in Figure 7. This figure indicates that the oligonucleotide Pso-GA₁₉-NLS is combined with the

glutathion beads, whereas the oligonucleotide Pso-GA₁₉ remains in the supernatant.

This clearly shows the capacity of the oligonucleotide Pso-GA₁₉-NLS to interact with the α -importin. The peptide part of the oligonucleotide-peptide chimeras is therefore indeed recognized by its receptor, which means that the peptide effectively fulfils its role of targeting signal.

10 Example 6

This example illustrates the possibility of coupling the oligonucleotide GA₁₉-SH to the peptide maleimide-NLS. Unlike Example 1, the chimera is not modified by a photoactivable alkylating agent.

15 The oligonucleotide GA₁₉-SH, of sequence 5'-AAGGAGAGGAGGGAGGGAA-3' (SEQ ID No. 4), with a thiol group at the 5' end, was coupled to the peptide maleimide-NLS which possesses a maleimide group at its N-terminal end under the same conditions as for the
20 oligonucleotide Pso-GA₁₉-SH (see Example 1).

The coupling is monitored by reversed-phase high-performance liquid chromatography (HPLC). It occurs with an equimolar stoichiometry: in two hours, the coupling is complete. The chimera is then purified
25 by HPLC.

The oligonucleotide-peptide chimera was

analysed by proteolytic action of trypsin as described in the "Materials and Methods" part.

The result is represented in Figure 8. It is comparable to that obtained with the oligonucleotide

5 Pso-GA₁₉-SH.

Example 7

This example illustrates the possibility of forming triple helices between the plasmid pXL2813 and

10 the GA-₁₉-NLS chimera in the absence of alkylating agent.

A kinetics of formation of triple helices between the plasmid pXK2813 and the GA-₁₉-NLS chimera, obtained as described in Example 5, was carried out and

15 studied according to the technique described in the "Materials and Methods" part, under the section "Formation of triple helices with the chimeras".

Figure 9 represents the kinetics of formation of the triple helices.

20 It appears that under the conditions of a plasmid concentration of 40 nM and an oligonucleotide concentration of 20 nM, a stable triple helix is formed.

25 Example 8

This example illustrates the characterization

of the peptide part of the GA₁₉-NLS chimera.

The peptide sequence used, the NLS signal of the SV40 T antigen, is recognized by receptors of the α -karyopherin family, as already mentioned in Example 4. The murine equivalent, called importin 60, fused with a glutathione S-transferase group, was used to characterize the oligonucleotide-peptide conjugates. It was performed as described in "Materials and Methods", under the section "Interactions with the importins".

The interactions between the glutathione-beads coated with importins 60 and the oligonucleotides GA₁₉ or GA₁₉-NLS were studied. After incubating for 30 minutes at room temperature, the pellet of beads (containing the components which interact with the importins) is separated from the supernatant.

The results are indicated in Figure 10. It appears that the oligonucleotide GA₁₉-NLS is combined with the glutathione beads, whereas the oligonucleotide GA₁₉ remains in the supernatant.

This clearly shows the capacity of the oligonucleotide GA₁₉-NLS to interact with the α -importin. The peptide part of the oligonucleotide-peptide chimeras is therefore indeed recognized by its receptor, and fulfils, here again, its role as targeting signal.

Example 9

This example illustrates the possibility of coupling the oligonucleotide pim-SH to the peptide maleimide-NLS.

5 The oligonucleotide pim-SH, of sequence
5'-GGGGAGGGGGAGG-3' (SEQ ID No. 15), with a thiol group
at the 5' end, was coupled to the peptide maleimide-NLS
which possesses a maleimide group at its N-terminal end
under the same conditions as for the oligonucleotide
10 GA₁₉-SH (see Example 5).

 The coupling is monitored by reversed-phase
high-performance liquid chromatography (HPLC). It
occurs with an equimolar stoichiometry: in two hours,
the coupling is complete. The chimera is then purified
15 by HPLC.

Example 10

This example illustrates the possibility of forming triple helices between the plasmid pXL2997 and
20 the pim-NLS chimera in the absence of alkylating agent.

 A kinetics of formation of triple helices
between the plasmid pXL2997 and the pim-NLS chimera
(formation obtained as described in Example 8), was
carried out and studied according to the technique
25 described in the "Materials and Methods" part, under
the section "Formation of triple helices with the

chimera".

Figure 12 represents the kinetics of formation of the triple helices.

It appears that under the conditions of
5 plasmid concentrations of 40 nM and of oligonucleotide concentrations of 20 nM, a stable triple helix is formed.

The invention having now been completely described, it is evident that numerous modifications
10 can be made to the present invention without as a result parting from the spirit of the invention and from the scope of the claims which follow.

CLAIMS

1. Vector for transferring nucleic acids,
characterized in that it comprises a double-stranded
5 DNA molecule and at least one oligonucleotide which is
coupled to a targeting signal and which is capable of
forming a triple helix with a specific sequence present
on the said double-stranded DNA molecule.

2. Vector for transferring nucleic acids
10 according to claim 1, characterized in that the double-
stranded DNA molecule is a plasmid or an episome.

3. Vector for transferring nucleic acids
according to claim 2, characterized in that the double-
stranded DNA molecule is a plasmid in circular form or
15 in a supercoiled state.

4. Vector for transferring nucleic acids
according to claims 1 to 3, characterized in that the
double-stranded DNA molecule comprises an expression
cassette consisting of one or more genes of interest
20 under the control of one or more promoters or of a
transcriptional terminator which is active in mammalian
cells.

5. Vector for transferring nucleic acids
according to claim 4, characterized in that the gene of
25 interest is a nucleic acid encoding a therapeutic
product.

6. Vector for transferring nucleic acids

according to one of claims 1 to 5, characterized in that the specific sequence present on the double-stranded DNA molecule is a homopurine-homopyrimidine sequence.

5 7. Vector for transferring nucleic acids according to one of claims 1 to 6, characterized in that the specific sequence present on the double-stranded DNA molecule is a sequence which is naturally present on the double-stranded DNA or a synthetic or a
10 natural sequence which is introduced artificially into the double-stranded DNA.

 8. Vector for transferring nucleic acids according to one of claims 1 to 7, characterized in that the oligonucleotide comprises a sequence poly-CTT
15 and the specific sequence present on the double-stranded DNA molecule is a sequence poly-GAA.

 9. Vector for transferring nucleic acids according to one of claims 1 to 7, characterized in that the oligonucleotide comprises the sequence
20 GAGGCTTCTTCTTCTTCTTCTT (SEQ ID No. 1) or the sequence (CTT)₇ (SEQ ID No. 2).

 10. Vector for transferring nucleic acids according to one of claims 1 to 7, characterized in that the specific sequence present on the double-
25 stranded DNA molecule comprises the sequence 5'-AAGGGAGGGAGGAGAGGAA-3' (SEQ ID No. 3) and the oligonucleotide comprises the sequence

5'-AAGGAGAGGAGGGAGGGAA-3' (SEQ ID No. 4) or
5'-TTGGTGTGGTGGGTGGGTT-3' (SEQ ID No. 5).

11. Vector for transferring nucleic acids
according to one of claims 1 to 7, characterized in
5 that the specific sequence present on the double-
stranded DNA molecule comprises all or part of the
sequence 5'-CTTCCCGAAGGAGAAAGG-3' (SEQ ID No. 6)
present in the replication origin ColE1 of E. coli, and
the oligonucleotide possesses the sequence
10 5'-GAAGGTTCTTCCCTCTTCC-3' (SEQ ID No. 7).

12. Vector for transferring nucleic acids
according to one of claims 1 to 7, characterized in
that the specific sequence present on the double-
stranded DNA molecule comprises the sequence
15 5'-GAAAAAGGAAGAG-3' (SEQ ID No. 8) or the sequence
5'-AAAAAAGGGAATAAGGG-3' (SEQ ID No. 10) present in the
 β -lactamase gene of the plasmid pBR322 and of E. Coli,
respectively.

13. Vector for transferring nucleic acids
20 according to one of claims 1 to 7, characterized in
that the specific sequence present on the double-
stranded DNA molecule comprises the sequence
5'-AAGAAAAAAGAA-3' (SEQ ID NO. 9) present in the
replication origin γ of the plasmids containing a
25 conditional replication origin such as pCOR.

14. Vector for transferring nucleic acids
according to one of claims 1 to 13, characterized in

that the oligonucleotide comprises at least 3 base pairs.

15. Vector for transferring nucleic acids according to claim 14, characterized in that the
5 oligonucleotide comprises 5 to 30 base pairs.

16. Vector for transferring nucleic acids according to one of claims 1 to 15, characterized in that the oligonucleotide exhibits at least one chemical modification making it resistant to, or protecting it
10 from, nucleases, or increasing its affinity towards the specific sequence present on the double-stranded DNA molecule.

17. Vector for transferring nucleic acids according to one of claims 1 to 16, characterized in
15 that the oligonucleotide is a succession of nucleosides which have undergone modification of the backbone.

18. Vector for transferring nucleic acids according to one of claims 1 to 16, characterized in that the oligonucleotide is coupled to an alkylating
20 agent forming a covalent bond at the level of the bases of the double-stranded DNA.

19. Vector for transferring nucleic acids according to claim 18, characterized in that the said alkylating agent is photoactivable.

25 20. Vector for transferring nucleic acids according to claims 18 and 19, characterized in that the said alkylating agent is a psoralen.

21. Vector for transferring nucleic acids according to claims 1 to 20, characterized in that the targeting signal interacts with a component of the extracellular matrix, a plasma membrane receptor, targets an intracellular compartment, and/or improves the intracellular flow of the double-stranded DNA.

22. Vector for transferring nucleic acids according to claim 21, characterized in that the said targeting signal comprises growth factors (EGF, PDGF, TGF β , NGF, IGF, I, FGF), cytokines (IL-1, IL-2, TNF, Interferon, CSF), hormones (insulin, growth hormone, prolactin, glucagon, thyroid hormone, steroid hormones), sugars which recognize lectins, immunoglobulins, transferrin, lipoproteins, vitamins such as vitamin B12, peptide or neuropeptide hormones (tachykinins, neurotensin, VIP, endothelin, CGRP, CCK, and the like), or any unit recognized by the integrins, for example the peptide RGD, or by other extrinsic proteins of the cell membrane.

23. Vector for transferring nucleic acids according to claim 21, characterized in that the targeting signal is an intracellular targeting signal, such as a nuclear homing sequence (NLS).

24. Vector for transferring nucleic acids according to claim 23, characterized in that the nuclear homing signal is the NLS sequence of the SV40 T antigen.

25. Vector for transferring nucleic acids according to claim 21, characterized in that the targeting signal allows both extracellular targeting and intracellular targeting.

5 26. Vector for transferring nucleic acids according to claims 1 to 25, characterized in that the coupling of the targeting signal to the oligonucleotide is obtained by synthesis on a solid phase or in solution, in particular by establishing disulphide,
10 thioether, ester, amide or amine bonds.

27. Composition, characterized in that it contains at least one vector as defined in one of claims 1 to 26.

15 28. Composition according to claim 27, characterized in that it contains, in addition, one or more transfecting agents.

29. Composition according to claim 28, characterized in that the transfecting agent is a cationic lipid, a lipopolyamine or a cationic polymer.

20 30. Composition according to claims 27 to 29, characterized in that it contains, in addition, one or more adjuvants capable of combining with the vector as defined in one of claims 1 to 24/transfecting agent complexes.

25 31. Composition according to claim 30, characterized in that the adjuvant is one or more neutral lipids chosen from natural or synthetic lipids

which are zwitterionic or which lack ionic charge under physiological conditions.

32. Composition according to either of claims 30 and 31, characterized in that the neutral lipid(s) is(are) chosen from lipids containing two fatty chains.

33. Composition according to claims 30 to 32, characterized in that the neutral lipid(s) is (are) chosen from dioleoylphosphatidylethanolamine (DOPE), oleoylpalmitoylphosphatidylethanolamine (POPE), di-stearoyl, -palmitoyl, -mirystoylphosphatidylethanolamines as well as their derivatives which are N-methylated 1 to 3 times, phosphatidylglycerols, diacylglycerols, glycosyldiacylglycerols, cerebrosides (such as in particular galactocerebrosides), sphingolipids (such as in particular sphingomyelins) or asialogangliosides (such as in particular asialoGM1 and GM2).

34. Composition according to claim 30, characterized in that the adjuvant is or comprises a compound which is involved in the condensation of the DNA.

35. Composition according to claim 34, characterized in that the said compound is derived, as a whole or in part, from a histon, a nucleolin and/or a protamine, or consists, as a whole or in part, of peptide units (KTPKKAKKP SEQ ID No.16) and/or (ATPAKKAA

SEQ ID No.17) repeated continuously or otherwise, it being possible for the number of units to vary between 2 and 10.

36. Composition according to any one of
5 claims 27 to 35, characterized in that it comprises a pharmaceutically acceptable vehicle for an injectable formulation.

37. Composition according to any one of
claims 27 to 35, characterized in that it comprises a
10 pharmaceutically acceptable vehicle for application to the skin and/or to the mucous membranes.

38. Use of a vector for transferring nucleic acids as defined in one of claims 1 to 26 for the manufacture of a medicament intended to treat diseases.

15 39. Method of transfecting nucleic acids into cells, characterized in that it comprises the following steps:

(1) synthesis of the oligonucleotide-targeting signal chimera,
20 (2) bringing the chimera synthesized in (1) into contact with a double-stranded DNA so as to form triple helices,

(3) optionally, complexing the vector obtained in (2) with one or more transfection agents
25 and/or one or more adjuvants, and

(4) bringing the cells into contact with the complex formed in (2) or, if applicable, in (3).

40. Method of treating diseases by
administration of a vector for transferring nucleic
acids as defined in any one of Claims 1 to 26
containing a double-stranded DNA capable of correcting
5 the said disease.

41. Recombinant cell containing a nucleic
acid transfer vector as defined in any one of claims 1
to 26.

42. Recombinant cell according to claim 41,
10 characterized in that it is a eukaryotic cell.

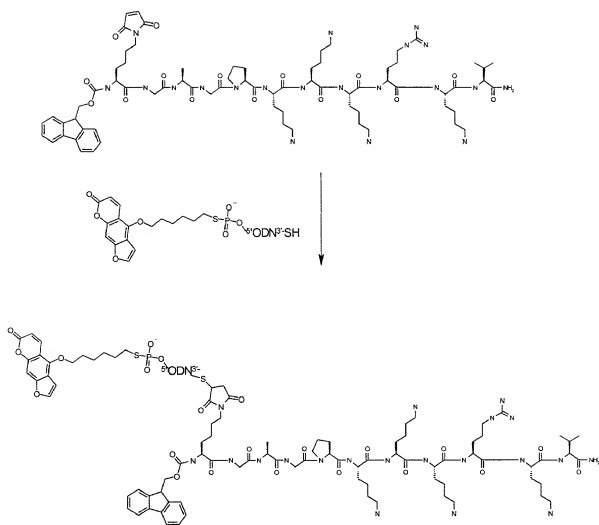
FIG. 1/13

FIG. 2/13

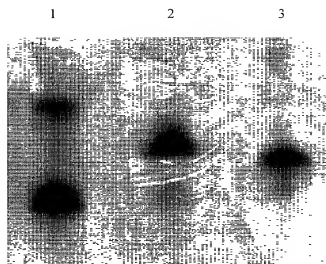


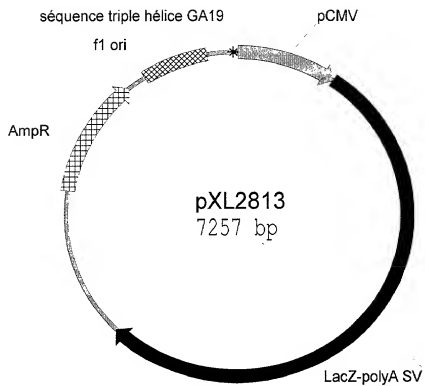
FIG. 3/13

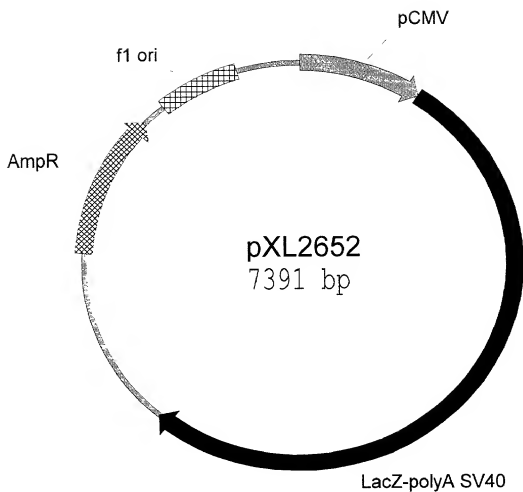
FIG. 4/13

FIG. 5/13

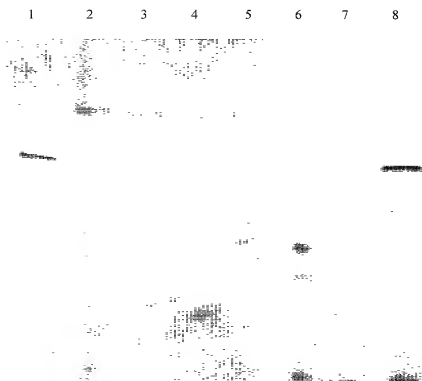


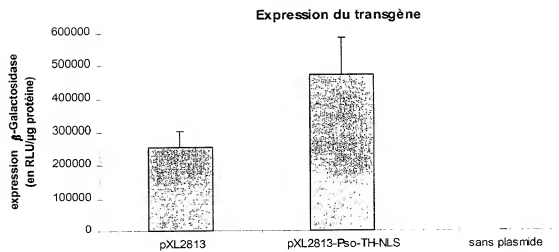
FIG. 6/13

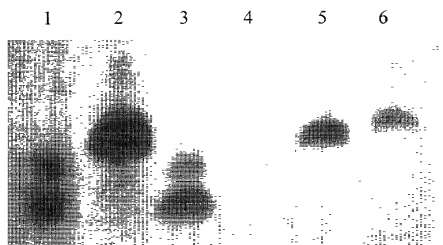
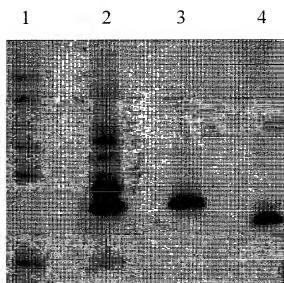
FIG. 7/13

FIG. 8/13

09/646399

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FIG. 9/13

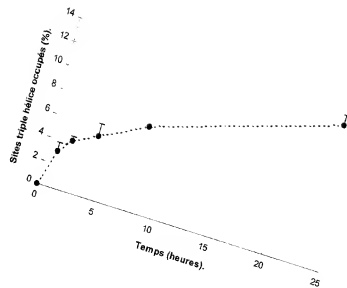


FIG. 10/13

1 2 3 4 5 6

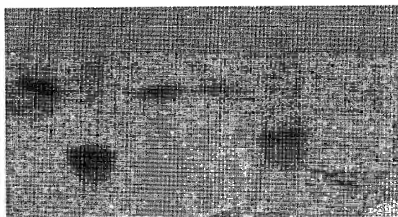


FIG. 11/13

Plasmide pXL2997

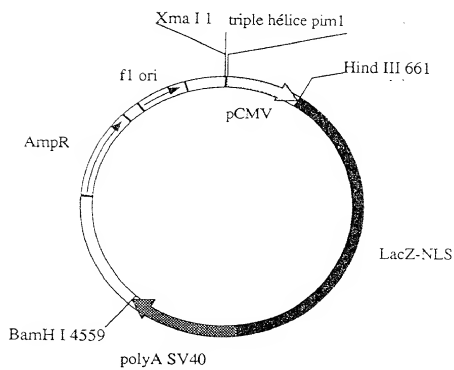
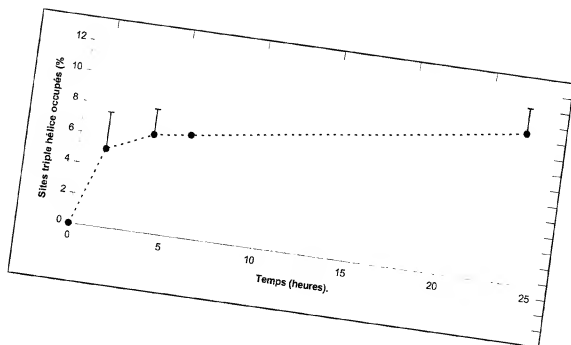


FIG. 12/13

1

Questions are asked about the following:



DECLARATION AND POWER OF ATTORNEY FOR PATENT APPLICATION

As a below named inventor(s), I/We hereby declare that:

My/Our residence(s), post office address(es) and citizenship(s) are as stated below my name(s).
I/We verily believe I am/we are the original, first and sole/joint inventor(s) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

VECTORS FOR TRANSFERRING NUCLEIC ACIDS, COMPOSITIONS CONTAINING THEM AND THEIR USES

and the specification of which (check one) ☐ is attached hereto (Attorney Docket No. _____) ☒ was filed on **September 15, 2000** as U.S. Application Number _____ and was amended on _____ (if applicable). ☐ was described and claimed in PCT Int'l Application Number _____ filed on _____ and as amended under PCT Article 19 on _____ (if any).

I/We hereby state that I/We have reviewed and understand the contents of the above identified specification, including the claims as amended by any amendment referred to above.
I/We acknowledge the duty to disclose to the U.S. Patent and Trademark Office all information known to me/us to be material to patentability as defined in 37 C.F.R. 1.56.

☒ I/We hereby claim foreign priority benefits under Title 35, United States Code §119(a)-(d) or 365 (b) of any foreign application(s) for patent or inventor's certificate, or 365(a) of any PCT international application which designated at least one other country other than the United States of America, listed below and having a filing date before that of the application on which priority is claimed. I/We have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

Foreign Priority:	FR98/03573 Number	France Country	24 March 1998 Day/Month/Year Filed
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Prior Foreign Appln(s):	PCT/FR99/00643 Number	PCT Country	19 March 1999 Day/Month/Year Filed
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☐ I/We hereby claim the benefit under Title 35, United States Code §119(e) of any United States Provisional application(s) listed below:

Number	Filing Date
--------	-------------

☐ I/We hereby claim the benefit under Title 35, United States Code §120 or 365(c) of any United States application(s) or international application designating the United States listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code §112, I/We acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations §1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application.

Application Serial No.	Filing Date	Status (Patented, Pending)
------------------------	-------------	----------------------------

I/We hereby appoint the attorneys and/or agents associated with the Customer No.(s) provided below as my/our attorneys and/or agents with full power to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith:

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I/We hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Title 18, United States code §1001, and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

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